

Combined effect of GSTM1, GSTT1 and GSTP1 polymorphisms on histological subtypes of lung cancer

R. C. SOBTI¹, PUSHPINDER KAUR¹, SATINDER KAUR¹, A. K. JANMEJA², S. K. JINDAL³, J. KISHAN⁴, & SARA RAIMONDI⁵

¹Department of Biotechnology, Panjab University, Chandigarh 160014, India, ²Department of Chest and Infectious Diseases, Government Medical College Hospital, Chandigarh 160032, India, ³Department of Pulmonary Medicine, PGIMER, Chandigarh 160012, India, ⁴TB and Chest Disease Hospital, Government Medical College Patiala, Punjab, India and ⁵Fondazione Policlinico IRCCS, Milan, Italy

Abstract

Genetic polymorphisms are natural genetic variations in the gene sequence that occur at a frequency of >1% in the population. This genetic variability (polymorphisms) can be a factor in cancer risk. The functional polymorphisms in GST genes play an important role in susceptibility to lung cancer. In our previous study, we reported that the combination of certain genotypes of GSTM1, GSTT1 and CYP1A1 is associated with lung cancer. The study has been extended to investigate the potential role of polymorphism in GSTP1 alone or in combination with the status of GSTM1 and GSTT1 genes in the likelihood of development of lung cancer. A total of 302 subjects (151 cases and 151 controls) were evaluated. Using a casecontrol design, individuals were genotyped for GSTs using multiplex polymerase chain reaction and restriction fragment length polymorphism techniques. The data obtained were analyzed using multiple logistic regression. The combined 'at risk' genotypes of GSTM1 null and GSTT1 null in comparison with 'wild-type' genotypes seems to be associated with a greater risk of lung cancer, but the results are not significant (odds ratio (OR) 2.0, 95% confidence interval (CI) 0.68-5.96) and for squamous cell carcinoma (SqCC) it was 1.6-fold (OR 1.6, 95% CI 0.49-5.68). In summary, our case-control study of lung cancer revealed that the effect of these polymorphisms is not very marked for different genotypic combinations of GSTP1, GSTM1 and GSTT1 in the context of developing lung cancer in a north Indian population. However, the increased risk was limited to SqCC, and was not found for other histological subtypes. Further analyses on this topic are needed.

Keywords: Genetic polymorphism, GSTs, lung cancer risk

(Received 7 June 2007; accepted 4 December 2007)

Introduction

Most common cancers such as those of lung, larynx, mouth, oesophagus, bladder and kidney result from the interaction between genetic and environmental factors (Bennet et al. 1999). Among the various factors, cigarette smoking plays an important role in

Correspondence: Professor R. C. Sobti, Department of Biotechnology, Panjab University, Chandigarh 160014, India. Tel: +91-172-2534087 (laboratory); +91-09417044523 (mobile). Fax: +91-172-2541409 (office). E-mail: rcsobti@pu.ac.in

ISSN 1354-750X print/ISSN 1366-5804 online © 2008 Informa UK Ltd.

DOI: 10.1080/13547500701843437



the aetiology of lung cancer. It is the most common cause of cancer-related deaths in both men and women. Lung cancer can be divided into two major histological groups - small cell lung carcinoma (SCLC) and non-small cell lung carcinoma (NSCLC) (Pass et al. 1996). Although studies evaluating the role of genetic polymorphisms in lung cancer risk have generally grouped all NSCLC together, histologically NSCLC includes the three most common subtypes – squamous cell carcinoma (SqCC), large cell carcinoma and adenocarcinoma (AC) (Ginsberg et al. 1997). Controversy still exists as to whether the different histological subtypes are different manifestations of the same disease process or different processes shared by some distinct risk factors. In order to demonstrate the important effect of environmental factors on the risk of lung cancer, a search for gene susceptibility was carried out.

Numerous polymorphisms occur in the genes encoding glutathione S-transferases (GSTs) (Mannervik 1985, Hayes & Pulford 1995). Among the GST enzymes, GSTM1, GSTT1 and GSTP1 play a major role in the detoxification of metabolites of carcinogens in tobacco smoke (Vos & Blanderen 1990). GSTM1 is expressed at high levels in the liver, but has also been found in several tissues including the lung (Mace et al. 1998, Strange et al. 1999). GSTM1 is known to detoxify arene oxides, including the ultimate carcinogenic form of benzo[a]pyrene (BP), BP-diol epoxide, where it is not involved in the detoxification of aromatic amines. The GSTT1 enzyme is expressed at high levels in the liver and in many extrahepatic tissues including the lung (Juronen et al. 1996). The GSTT1 null phenotype (GSTT1*0/*0) is a result of a deletion of the entire GSTT1 gene (Pemble et al. 1994). GSTP1 is widely expressed in different human epithelial tissues (Terrier et al. 1990) and is the major GST in human lungs (Antilla et al. 1993). The polymorphism of GSTP1 is associated with an increased risk of lung, bladder and testicular cancers (Harries et al. 1997, Watson et al. 1998). The GSTP1 gene is polymorphic with respect to a single base change in exons 5 and 6, which results in an amino acid change from Ile/ Val and Ala/Val, respectively, leading to reduced enzyme activity (Board et al. 1989, Johansson et al. 1998). The enzyme activity is affected by substitution at position 105, which is located in a hydrophobic substrate-binding site. Individuals with the 105 Val allele seem to have a higher risk of developing cancer than those with the 105 Ile allele, as the former allele has a higher catalytic efficiency than the latter for carcinogenic aromatic epoxides (Sundberg et al. 1998). Many studies have examined the association between GSTP1 polymorphisms and risk of lung cancer, but no statistically significant associations have been obtained (Mironova et al. 1998, Kihara et al. 1999, To-Figureas et al. 2001, Lewis et al. 2002, Wang et al. 2003). GSTM1 has been studied with respect to polymorphisms and cancer risk. It detoxifies active metabolites of polycyclic aromatic hydrocarbons (PAHs) (Hayes et al. 1995). GSTT1 is also known to be involved in the detoxification of several environmental carcinogens such as 1,3-butadiene and ethylene oxide in tobacco smoke and ambient air (Landi 2000) and it has been documented that GSTM1 and GSTT1 genes are missing in some individuals. The association of null genotypes of GSTM1 and GSTT1 has been widely studied, but results with respect to their association with risk of lung cancer are contradictory (Seideigard et al. 1990, D'Errico et al. 1996, Ryberg et al. 1997, LeMarchand et al. 1998, Mironova et al. 1998, Kiyohara et al. 2000, Stucker et al. 2002).

Individuals who have more than one risk-associated polymorphism may have a greater risk of developing lung cancer. Some studies have reported that the



combination of GSTP1 (Val/Val) and GSTM1 null genotypes is associated with a higher risk of lung cancer (Kihara et al. 1999, Mironova et al. 1998, Wang et al. 2003). In our earlier study, we reported that the combination of GSTM1, GSTT1 with CYP1A1 genotypes promotes a higher risk of lung cancer (Sobti et al. 2004). Another study, however, has found that lung cancer risk is not associated with GSTT1 and GSTP1 (Lewis et al. 2002).

We have extended our investigations for checking the potential role of GSTP1 gene polymorphisms alone and in combination with GSTM1 and GSTT1 genes on susceptibility to lung cancer in a north Indian population.

Materials and methods

Sample collection

Histologically confirmed incident adult patients with lung cancer (n = 151) reviewed by a pathologist were recruited from three medical institutes - the Post Graduate Institute of Medical Education and Research (PGIMER), the Government Medical College Hospital (GMCH) both at Chandigarh, India and the TB and Chest Diseases Hospital of Government Medical College, Patiala (Punjab, India) of north India. None of the patients had received radiation or chemotherapy. The relevant demographic and epidemiological information including detailed diet and smoking information along with informed consent was obtained through a questionnaire. The controls (n=151) were also hospital based and were free of benign and malignant tumours both at the time of, and prior to, diagnosis, but were admitted for minor pulmonary complaints such as respiratory infection and bronchitis. Peripheral blood samples were obtained from all subjects and DNA was extracted using the SDS/ proteinase K and phenol-chloroform method (http://www.genome.ou.edu/protocols_ book/protocols_partIII.html).

Histological data were available for 143 cases, whereas no such information could be obtained for the remaining eight cases (non-classified). The tumours were not defined, as they were so advanced that histological diagnosis was not relevant to the palliative treatment given. This study was approved by the human subjects ethical committees of all the involved institutions.

Genotyping of GSTM1 and GSTT1 genes

The GSTM1 and GSTT1 genetic polymorphisms were evaluated using multiplex polymerase chain reaction (PCR) techniques as already described (Arand et al. 1996). The primer sequences (Sigma Aldrich, St Louis, MO, USA) are shown in Table I.

Table I. The primer sequences for GSTM1 and GSTT1 and control albumin.

| Genes | Primer sequence | Size (bp) | |
|---------|------------------------------|-----------|--|
| GSTM1 | (F)-GAACTCCCTGAAAAGCTAAAGC | 215 | |
| | (R)-GTTGGGCTCAAATATACGGTGG | | |
| GSTT1 | (F)-TTCCTTACTGGTCCTCACATCTC | 480 | |
| | (R)-TCACCGGATCATGGCCAGCA | | |
| Albumin | (F)-GCCCTCTGCTAACAAGTCCTAC | 350 | |
| | (R)-GCCCTAAAAAGAAAATCGCCAATC | | |



Each PCR reaction mixture (50 µl) contained 10 pmol of each primer, 2.5 mM of MgCl₂, 0.2 mM each deoxynucleotide triphosphate (dNTPs), 1 unit of Taq polymerase, 100 ng of genomic DNA and 5% DMSO. Amplification was performed with initial denaturation at 94°C for 5 min, followed by 30 cycles of denaturation at 94°C, annealing at 64°C and extension at 72°C for 1 min each and a final extension at 72°C for 7 min. DNA fragments of expected size were amplified using primers specific for GSTT1 and GSTM1 genes, respectively (Figure 1). The albumin gene was used as internal standard. To detect the pattern of genes, the amplified DNA fragments were electrophoresed through a 2% agarose gel and observed under UV light after staining with ethidium bromide.

The GSTP1 gene in exon 5 at codon 105 was analyzed by the method of Kihara et al. (1999). The DNA was amplified with the GSTP1 primers (F) - GTAGTTTGCCCAAGGTCAAG, (R) - AGCCACCTGAGGGGTAAG (Sigma Aldrich). PCR reaction mixture (50 µl) was prepared as described above except for the addition of DMSO. The cycling parameters included initial denaturation at 94°C for 5 min, followed by 30 cycles of denaturation at 94°C for 1 min, annealing at 59°C for 1 min 30 s, and extension at 72°C for 1 min 30 s. The final polymerization step was carried out at 72°C for 7 min to complete the elongation process. PCR products were digested with 2 units of restriction enzyme Alw 261 (MBI Fermentas, USA) overnight at 37°C. The digestion distinguishes between the restriction sites on the Ile and Val alleles (ACa/gTCT), where small characters a/g represent the polymorphic site.

PCR-restriction fragment length polymorphism (RFLP) patterns resulted in a band of 113 bp in all samples, which represented a control cut for confirmation of proper digestion. In the wild-type GSTP1 (Ile/Ile), bands of 329 and 113 bp were generated,

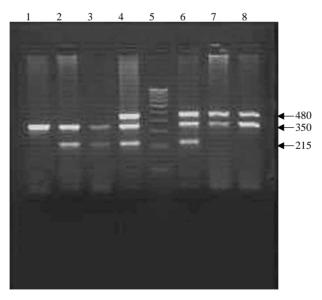


Figure 1. Polymerase chain reaction amplified product of GSTM1T1. Lane 1, GSTM1T1 null; lanes 2 and 3, GSTM1 (present) (215 bp), GSTT1 (null); lanes 4 and 6, GSTM1 (present) (215 bp), GSTT1 (present) (480bp); lanes 7 and 8 GSTT1 (present) (480 bp), GSTM1 (null); lane 5, 100 bp DNA marker.



whereas in the homozygous mutant type GSTP1 (Val/Val), the bands of 216, 113 and 107 bp were produced. In heterozygous mutant type GSTP1 (Ile/Val), all the four bands were present (Figure 2).

Statistical analysis

Age, gender, smoking status and genotypes of GSTM1, GSTT1 and GSTP1 genes were tabulated for cases and controls. To determine any difference between cases and controls according to age, number of cigarettes smoked per day, pack-years and gender, t-tests and γ^2 tests were performed. Cases were further categorized into various histological subtypes, in order to find out any specific association of genetic polymorphisms according to histological type.

The association between polymorphisms in GST genes and risk of lung cancer was estimated by computing odds ratios (ORs) and 95% confidence interval (CI) using a multivariate logistic regression analysis which included several potential confounding variables (e.g. age, gender and smoking status - smokers and people who had never smoked, separately). Subjects were considered as non-smokers if they had never smoked as much as one cigarette per day for a year, while the smokers were those who had smoked cigarettes over their lifetime. Under the hypothesis that GSTM1, GSTT1 and GSTP1 genotypes may be differentially associated with histological subtypes, the ORs for each polymorphism and for the combinations between GSTP1 and either of the GST types (GSTM1 or GSTT1) were tested in the models also for SCLC, NSCLC, SqCC and AC. Statistical analysis was performed using SAS version 8.

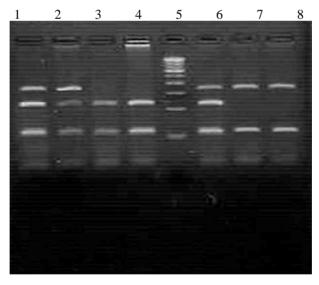


Figure 2. Restriction fragment length polymorphism analysis of GSTP1. Lanes 1, 2 and 6, heterozygous mutant (329, 216, 113 and 107 bp); lanes 3 and 4, homozygous mutant (216, 113 and 107 bp); lanes 7 and 8, homozygous wild (329 and 113 bp); lane 5, 100 bp DNA marker.



Results

Baseline characteristics

Demographic variables such as age, gender, and smoking status have been summarized in Table II, according to the case/control status. The average age was calculated as 56.9 ± 10.4 years for cases and 56.4 ± 11.1 years for controls (p = 0.67). Most of the study participants were smokers with the higher percentage of these among cases (81.4%) than controls (74.1%). Moreover, the cancer cases had a higher value of pack years (64.1 ± 50.8) than controls (37.9 ± 29.2) (p < 0.0001), while the average number of cigarettes smoked per day was 14.4 ± 12.2 for cases and 8.7 ± 8.8 for controls (p < 0.0001). The frequencies of males and females in the case and control groups were comparable (p = 0.27).

Distribution of GSTM1, GSTT1 and GSTP1 polymorphism in north Indian population

The frequency of GSTM1 and GSTT1 null genotypes was higher in cases (41.7% and 17.8%, respectively) compared with the controls (35% and 13.2%, respectively). The frequencies of the three genotypes of GSTP1 were as follows: Ile/Ile 51.6%, Ile/Val 45% and Val/Val 3.3% in cases and Ile/Ile 41%, Ile/Val 55.0% and Val/Val 4.0% in controls. Table II also presents the association of polymorphisms in individual GST genes with lung cancer. The GSTM1 (null) genotype was not found to be a risk factor for developing lung carcinoma compared with the controls. But the GSTT1 (null) genotype was found to be a marginal risk (OR 1.4, 95% CI 0.74–2.62). It seems that GSTP1 (Ile/Val) could be a protective factor for the development of lung cancer, even if the association is borderline (OR 0.63, 95% CI 0.40-1.01).

Table II. Descriptive characteristics of the study population.

| Variables | Cases | Control | OR (95% CI) | |
|-------------------------|-----------------|-----------------|------------------|--|
| Sample size, n | 151 | 151 | | |
| Age (years), mean ±SD | 56.9 ± 10.4 | 56.4 ± 11.1 | | |
| Gender, n (%) | | | | |
| Male | 130 (86.0) | 123 (81.4) | | |
| Female | 21 (13.9) | 28 (18.5) | | |
| Smoking status, n (%) | | | | |
| Non-smoker ^a | 28 (18.5) | 39 (25.8) | 1 (ref.) | |
| Smokers ^b | 123 (81.4) | 112 (74.1) | 1.48 (0.85–2.56) | |
| GSTM1 (%) | | | | |
| Present | 88 | 98 | 1 (ref.) | |
| Null | 63 | 53 | 1.30 (0.81–2.08) | |
| GSTT1 (%) | | | | |
| Present | 124 | 131 | 1 (ref.) | |
| Null | 27 | 20 | 1.39 (0.74–2.62) | |
| GSTP1 (%) | | | | |
| Ile/Ile | 78 | 62 | 1 (ref.) | |
| Ile/Val | 68 | 83 | 0.63 (0.40-1.01) | |
| Val/Val | 5 | 6 | 0.64 (0.19-2.22) | |

OR, odds ratio adjusted for age and gender; CI, confidence interval; ref., reference.



^aNon-smokers (never smoked as much as one cigarette per day for a year); ^bsmokers (smoked cigarettes over their lifetime).

Distribution of GST polymorphism in histological subtypes

Further, the variables were categorized for lung cancer histological subtypes as SCLC, NSCLC, SqCC, AC and NC. Among 151 cases in the present study, 63.5% were identified to have SqCC, 15.8% SCLC, 7.9% NSCLC, 7.2% AC and 5.2% NC (Table III).

When genotypes of GSTs were considered with the risk to individual subtypes, a 2.4-fold increased risk in AC was found with the GSTM1 gene, although the result was not significant. Patients with NSCLC also showed an OR higher than 1.00 (1.77, 95% CI 0.54-5.84). A borderline increased risk was observed (OR 3.76, 95% CI 0.99-14.28) for NSCLC that presented a deletion in the GSTT1 gene. The individuals with GSTP1 (Ile/Val) were at marginal increased risk of AC subtype (OR 1.30, 95% CI 0.36–4.66) compared with controls. No significant correlation was observed between polymorphisms in GSTs and their association with smoking and drinking habits.

Gene-gene interactions

GSTM1-GSTT1 genotypes. Individuals having both GSTM1 and GSTT1 null genotypes had an increased risk for lung cancer (OR 2.0, 95% CI 0.68-5.96) while for SqCC it was 1.6-fold (OR 1.6, 95% CI 0.49-5.68) (Table IV).

GSTM1-GSTP1 genotypes. The ORs and 95% CI were also calculated for lung cancer risk in relation to both GSTM1 and GSTP1 genotypes and there was a weak association in all the GSTM1 and GSTP1 genotypes (Table V). Individuals with GSTM1 null and GSTP1 (Ile/Ile) showed increased risk towards NSCLC (OR 2.5, 95% CI 0.45-14.05) and AC (OR 1.7, 95% CI 0.26-11.64). Similarly, with the GSTM1 (null) and GSTP1 (Ile/Val/Val) combined genotype, an association was observed with AC (OR 2.4, 95% CI 0.47-12.34) and NSCLC (OR 1.8, 95% CI 0.33-10.46) development.

Table III. Odds ratio^a (95% confidence interval) for GSTM1, GSTT1 and GSTP1 polymorphisms stratified according with histological subtypes.

| Gene polymorphism | SqCC (n=96) (63.5%) | SCLC (n = 24) (15.8%) | NSCLC (n=12) (7.9%) | AC $(n=11)$ (7.2%) |
|----------------------|------------------------|--------------------------|------------------------|-----------------------|
| GSTM1 | | | | |
| Present | 1 (ref.) | 1 (ref.) | 1 (ref.) | 1 (ref.) |
| Null | 1.13 (0.66–1.93) | 1.29 (0.53–3.15) | 1.77 (0.54–5.84) | 2.41 (0.69-8.41) |
| GSTT1 | | | | |
| Present | 1 (ref.) | 1 (ref.) | 1 (ref.) | 1 (ref.) |
| Null | 1.19 (0.57–2.48) | 0.86 (0.23–3.21) | 3.76 (0.99–14.28) | 1.35 (0.27–6.85) |
| GSTP1 | | | | |
| Ile/Ile | 1 (ref.) | 1 (ref.) | 1 (ref.) | 1 (ref.) |
| Ile/Val | 0.59 (0.35–1.00) | 0.36 (0.14-0.92) | 1.07 (0.32–3.61) | 1.30 (0.36-4.66) |
| Val/Val | 0.59 (0.14–2.50) | 0.58 (0.06–5.34) | = | = , |

SqCC, squamous cell carcinoma; SCLC, small cell lung carcinoma; NSCLC, non-small cell lung carcinoma; AC, adenocarcinoma; ref., reference.



^aOdds ratio adjusted for age, gender and smoking status.

Table IV. Odds ratios^a (OR) for the combined effect of GSTM1 and GSTT1 genotype and risk of lung cancer.

| GSTM1 | GSTT1 | All OR | SqCC | SCLC | NSCLC | AC |
|---------|---------|----------------------------|----------------------------|---------------------------|----------------------------|----------------------------|
| Present | Present | 1.0 (ref.) (73/85) | 1 (51/85) | 1 (10/85) | 1 (4/85) | 1 (4/85) |
| Null | Null | 2.0 (0.68–5.96) (12/7) | 1.6 (0.49–5.68) (7/7) | 1.0 (0.15–6.81) (1/7) | 4.9 (1.05–23.36) (2/7) | 2.7 (0.35–22.03) (1/7) |
| Present | Null | 1.3 (0.56–3.24) (15/13) | 1.0 (0.36–2.88) (8/13) | 1.0 (0.27–4.35) (2/13) | 2.9 (0.59–13.21) (2/13) | 1.6 (0.19–13.21) (1/13) |
| Null | Present | 1.3 (0.75–2.21) (51/46) | 1.0 (0.59–2.01) (30/46) | 1.4 (0.49–3.86) (9/46) | 1.7 (0.47–6.81) (4/46) | 2.1 (0.61–7.76) (5/46) |

SqCC, squamous cell carcinoma; SCLC, small cell lung carcinoma; NSCLC, non-small cell lung carcinoma; AC, adenocarcinoma; ref., reference.

GSTT1-GSTP1 genotypes. The impact of combinations of genotypes of GSTT1 and GSTP1 on the risk of lung cancer was also evaluated. No significant correlation was observed between polymorphisms in GSTT1 and GSTP1 and their association with lung cancer risk (Table VI). When these data were dissected according to histological subtypes, the combination of GSTM1 (null) and GSTP1 (Ile/Ile) genotypes revealed 2.1-fold (95% CI 0.77-6.05) increased risk for SCLC, 2.5-fold (95% CI 0.34-19.25) for NSCLC and 3.3-fold (95% CI 0.41-27.27) for AC. In NSCLC, the combination of GSTT1 (null) with GSTP1 combined (Ile/Val/Val) genotype showed a 2.5-fold increased risk (95% CI 0.63–10.32). The risk came down to 1.5-fold with GSTM1 (present) and GSTP1 combined (Ile/Val/Val/Val).

Discussion

The most common form of human genetic variation is the single nucleotide polymorphism. Genetic polymorphism may contribute to individual susceptibility to cancer, but the molecular mechanism behind this is still not clear. In this case-control study, we are reporting for the first time the data on the association of GSTP1 with GSTM1 and GSTT1 in lung cancer as a whole and in various histological subtypes in a

Table V. Odds ratios (OR) for the combined effect of GSTM1 and GSTP1 genotype and risk of lung cancer.

| GSTM1 | GSTP1 | All OR | SqCC | SCLC | NSCLC | AC |
|---------|---------------------|-----------------------------|----------------------------|---------------------------|----------------------------|----------------------------|
| Present | Ile/Ile | 1.0 (ref.) (50/40) | 1 (34/40) | 1 (9/40) | 1 (2/40) | 1 (2/40) |
| Null | Ile/Ile | 1.0 (0.48–2.17) (28/22) | 0.9 (0.39–2.13) (17/22) | 1.2 (0.33–4.41) (6/22) | 2.5 (0.45–14.05) (3/22) | 1.7 (0.26–11.64) (2/22) |
| Null | Ile/Val/ Val/Val | 0.9 (0.45–1.80) | 0.7 (0.34–1.67) | 0.6 (0.21–1.86) | 1.8 (0.33–10.46) | 2.4 (0.47–12.34) |
| | | (35/31) | (20/31) | (4/31) | (3/31) | (4/31) |
| Present | Ile/Val/ Val/Val | 0.7 (0.52–0.97) p < 0.04 | 0.5 (0.25–1.03) | 0.4 (0.15–1.21) | 1.3 (0.26–7.07) | 1.0 (0.18–5.92) |
| | | (38/58) | (25/58) | (5/58) | (4/58) | (3/58) |

SqCC, squamous cell carcinoma; SCLC, small cell lung carcinoma; NSCLC, non-small cell lung carcinoma; AC, adenocarcinoma; ref., reference.



^aOR adjusted for age, gender and smoking status

^aOR adjusted for age, gender and smoking status. Significant p-value <0.05.

Table VI. Odds ratios (OR)^a for the combined effect of GSTT1 and GSTP1 genotype and risk of lung cancer.

| GSTT1 | GSTP1 | All OR | SqCC | SCLC | NSCLC | AC |
|---------|---------------------|---------------------------|--------------------------|--------------------------|---------------------------|---------------------------|
| Present | Ile/Ile | 1.0 (ref.) (66/57) | 1 (45/57) | 1 (12/57) | 1 (66/57) | 1 (3/57) |
| Null | Ile/Ile | 1.3 (0.93–1.86) (12/5) | 1.2 (0.69–2.21) (6/5) | 2.1 (0.77–6.05) (3/5) | 2.5 (0.34–19.25) (1/5) | 3.3 (0.41–21.27) (1/5) |
| Present | Ile/Val/ Val/Val | 0.6 (0.40–1.14) | 0.6 (0.34–1.12) | 0.5 (0.21–1.60) | 0.7 (0.20–3.00) | 1.5 (0.39–5.76) |
| | | (58/74) | (36/74) | (9/74) | (4/74) | (6/74) |
| Null | Ile/Val/ Val/Val | 0.8 (0.36–2.06) | 0.7 (0.28–2.06) | - | 2.5 (0.63–10.32) | 1.2 (0.14–11.22) |
| | | (15/15) | (9/15) | = | (3/15) | (1/15) |

SqCC, squamous cell carcinoma; SCLC, small cell lung carcinoma; NSCLC, non-small cell lung carcinoma; AC, adenocarcinoma; ref., reference.

north Indian population. In our lung cancer samples, there were no cases of the large cell subtype. All other lung histological subtypes were included in the main association analysis of overall lung cancer risk, but the number of NSCLC (n=12) and adenocarcinoma (n = 11) was too small. It was found that there is an association of GSTM1 and GSTT1 null genotypes with an increased risk of SqCC with lung cancer in general.

The prevalence of GST genotypes differs within India. The genotype frequencies in our control group were similar to those reported by other authors in the north Indian population (Mishra et al. 2004). In south India, 30.4% of the population lacked the GSTM1 gene and 16.8% lacked the GSTT1 gene. The highest frequency of GSTM1 null was observed in Karnataka (36.4%), while Andhra Pradesh had the lowest frequency of GSTM1 and GSTT1 combined null genotype (1.7%) (Naveen et al. 2004). In a study carried out in a north Indian population on the prostate (Mittal et al. 2004, Srivastava et al. 2005), cervix (Sharma et al. 2004) and oral cancer (Sreelekha et al. 2001), no significant association between the GSTM1 and GSTT1 null genotypes was found. Similarly for GSTP1, a non-significant association was observed by Mittal et al. (2005) in bladder cancer in a north Indian population. But Srivastava et al. (2004) found a significant association with the null alleles of GSTM1 and GSTT1 genotypes.

Many studies have tried to establish links between polymorphic expression of different GSTs and risk of lung cancer in different ethnic groups (Seideigard et al. 1990, Ryberg et al. 1997, Harries et al. 1997, Mironova et al. 1998, Kihara et al. 1999), but the results are conflicting (Lewis et al. 2002, Wang et al. 2003). Interindividual variability in GST enzyme activity is believed to confer differences in susceptibility to cancers with major environmental determinants such as lung cancer (Kato et al. 1995, Rebbeck 1997, Hirvonen et al. 1999). The inconsistency in results might be due to polymorphism frequency differences across different ethnicities. All the three genes GSTP1, GSTM1 and GSTT1 play an important role in the cellular defence system (Antilla et al. 1995, Mainwaring et al. 1996, Lewis et al. 2002, Wang et al. 2003). GSTP1 is widely expressed in normal human epithelial tissues and has been shown to be overexpressed in lung cancer (Terrier et al. 1990, Antilla et al. 1993). Two genetic polymorphisms are known for GSTP1. These are Ile-105-Val,



^aOR adjusted for age, gender and smoking status.

resulting from an A>G transition at base 1578, and Ala-114-Val, resulting from a C > T transition at base 2293. Two studies have examined the frequency of the Ile-105-Val GSTP1 and Ala-114-Val GSTP1 polymorphism in different cancers (Harries et al. 1997, Watson et al. 1998). Epidemiological and functional studies on the polymorphism of GSTP1 suggest that the Ile and Val alleles confer increased susceptibilities to different types of cancer (Sundberg et al. 1998).

In our study, an association between the polymorphisms of GSTs was observed in individuals with SqCC. It is possible that individuals with GSTM1 and GSTT1 null genotypes are more susceptible to carcinogenic substrates. In these a close relationship between carcinogen exposure and histological subtypes has been evaluated. Several studies have associated GSTP1 polymorphism at codon 105 with GSTM1 and GSTT1 with susceptibility to SCLC and AC (Alexandrie et al. 1994, Garcia-Closas et al. 1997, LeMarchand et al. 1998, Ford et al. 2000). Studies on different populations have also indicated that separate GSTM1, GSTT1 and GSTP1 polymorphisms are not statistically related to lung cancer, but there is a strong association between the combined GSTM1, GSTT1 and GSTP1 genotypes which interact to increase the risk of lung cancer (Table VII).

Investigations on the association between GSTP1 polymorphism and lung cancer have shown that the GSTP1 Val genotype is not significantly associated with lung cancer risk (Hayashi et al. 1992, Alexandrie et al. 1995, Kihara et al. 1995, Garcia-Closas et al. 1997, Ryberg et al. 1997, Watson et al. 1998, Mironova et al. 1998, Kihara et al. 1999, Ford et al. 2000, To-Figureas et al. 2001, Dialyna et al. 2003). However, Stucker et al. (2002) reported that genotypes deficient for GSTMI and GSTP1 are important risk modifiers for lung cancer. Cote et al. (2005) observed that specific combinations of GST polymorphism increased the risk of early-onset lung cancer. On the other hand, a meta-analysis carried out by Zheng et al. (2006) on 130 genetic association studies provides the most comprehensive assessment in relation to lung cancer of five GST gene polymorphisms which indicates that the risk of lung cancer is not strongly associated with I105V and A114V polymorphisms in the GSTP1 gene or with the GSTM3 intron 6 polymorphism. Similarly, Skuladottir et al. (2005) carried out a pooled study on lung cancer patients in Norway and Denmark

Table VII. Studies in different populations on GSTM1, GSTT1 and GSTP1 polymorphisms.

| | Significantly associated/ | | | | |
|--------------|---------------------------|------------------------------|------------------------|--|--|
| | Population | not significantly associated | Reference | | |
| GSTM1/GSTT1 | | | | | |
| Null/null | Greece | Significantly associated | Dialyna et al. 2003 | | |
| Null/null | Japanese | Significantly associated | Kiyohara et al. 2000, | | |
| | | | Hayashi et al. 1992, | | |
| | | | Kihara et al. 1995 | | |
| GSTM1/GSTP1 | | | | | |
| Null/Val/Val | Chinese | Significantly associated | Wang et al. 2003 | | |
| Null/Val/Val | Japanese | Significantly associated | Alexandrie et al. 1994 | | |
| GSTM1 | | | | | |
| Null | Chinese | Not significantly associated | Wang et al. 2003 | | |
| GSTP1 | | | | | |
| Val/Val | Chinese | Not significantly associated | Wang et al. 2003 | | |
| Smoking+GSTs | Chinese | Not significantly associated | Wang et al. 2003 | | |



populations on xenobiotic gene polymorphism and found a non-significant association with lung cancer. As in studies on Chinese and German populations (Miller et al. 2002, Wang et al. 2003), we have also not observed a significant effect of GSTP1 (Ile/ Val or Val/Val) genotype on lung cancer. The combination of GSTP1 (Ile/Val or Val/ Val) polymorphism with GSTM1 and GSTT1 null does not seem to be associated with increased risk of lung cancer. In our study, the control population consisted of a group of patients with a range of different diagnoses. GST polymorphism may be associated with altered risk in these conditions so that the risk in the case population may be biased; controls with no history of lung disease would be more suitable, but more difficult to recruit from a bronchoscopy clinic. After adjustment for age, gender and smoking status, our results still agree with the published studies on north Indian populations and other ethnic groups worldwide. Besides this, our study has several important strengths such as: (1) both cases and controls underwent the same diagnostic procedure, (2) the controls and cases were drawn from same base (ethnic) population, and (3) interviewing was blind to case status.

We have conducted this case-control study to evaluate the association between this polymorphism with specific histological subtypes and risk of lung cancer. The data thus obtained confirm results from other studies that Ile/Val or Val/ Val genotypes of GSTP1 polymorphism in combination with both GSTM1 and GSTT1 null genotypes do not have a marked tendency to develop lung cancer but show an increased risk for SqCC. Further investigations need to be performed with more samples on a histological basis.

Acknowledgements

The authors wish to thank all the staff, and the patients who took part in this study, at the Bronchoscopy Unit. This work was funded by Indian Council of Medical Research, New Delhi, India in the form of project to R. C. Sobti.

References

- Alexandrie AK, Sundberg MI, Seidegard J, Toruling G, Rannug A. 1994. Genetic susceptibility to lung cancer with special emphasis on CYP1A1 and GSTM1: a study on host factors in relation to age at onset, gender and histological cancer types. Carcinogenesis 15:1785-1790.
- Antilla S, Hirvonen S, Vainio H, Husgafvel-Pursiainen K, Hayes JD, Ketterer B. 1993. Immunohistochemical localization of glutathione S-transferases in human lung cancer. Cancer Research 53:5643-5648.
- Antilla A, Luostarinen L, Hirvonen A, Elovaara E, Karjalainen A, Nurminen T, Hayes JD, Vainio H, Ketterer B. 1995. Pulmonary expression of glutathione S-transferase M3 in lung cancer patients: association with GSTM1 polymorphism, smoking, and asbestos exposure. Cancer Research 55:3305-3309.
- Arand M, Muhlbauer R, Hengstler J, Jager E, Fuchs J, Winkler L, Oesch F. 1996. A multiplex polymerase chain reaction protocol for the simultaneous analysis of the glutathione-S-transferase GSTM1 and GSTT1 polymorphism. Annals of Biochemistry 236:184–186.
- Bennet WP, Alavanja MCR, Blomeke B, Vahakangas KH, Castren K, Welsh JA, Bowman ED, Khan MA, Flieder DB, Harris CC. 1999. Environmental tobacco smoke, genetic susceptibility, and risk of lung cancer in never smoking women. Journal of the National Cancer Institute 91:2009-2014.
- Board PG, Webb GC, Coggan M. 1989. Isolation of a cDNA clone and localization of the human glutathione S-transferase 3 genes to chromosome bands 11q13 and 12q13-14. Annals of Human Genetics 53:205-213.



- Cote ML, Kardia SLR, Wenzlaff AS, Land SJ, Schwartz AG. 2005. Combinations of glutathione S-transferase genotypes and risk of early-onset lung cancer in Caucasians and African Americans: a population-based study. Carcinogenesis 26:811-819.
- D'Errico A, Taioli E, Chen X, Vineis P. 1996. Genetic metabolic polymorphisms and the risk of cancer: a review of the literature. Biomarkers 1:149-173.
- Dialyna IA, Miyakis S, Georgatou N, Spandidos DA. 2003. Genetic polymorphisms of CYP1A1, GSTM1 and GSTT1 genes and lung cancer risk. Oncology Report 10:1829-1835.
- Ford JG, Li Y, O'Sullivan MM, Demopoulos R, Garte S, Taioli E, Brandt-Rauf PW. 2000. Glutathione S-transferase M1 polymorphism and lung cancer risk in African-American. Carcinogenesis 21:1971-
- Garcia-Closas M, Kelsey KT, Wiencke JK, Xu X, Wain JC, Christiani DC. 1997. A case control study of cytochrome P4501A1, glutathione S-transferase M1, cigarette smoking and lung cancer occur susceptibility (Massachusetts, United States). Cancer Causes & Control 8:544-553.
- Ginsberg RJ, Vokes EE, Raben A. 1997. Non-small cell lung cancer. In: DeVita VT Jr, Hellman S, Rosenberg SA, editors. Cancer: Principles and Practice of Oncology. Philadelphia, PA: Lippincott-Raven. p. 858-911.
- Harries LW, Stubbins MJ, Forman D, Howard GCW, Wolf CR. 1997. Identification of genetic polymorphisms at the glutathione S-transferase Pi Locus and association with susceptibility to bladder, testicular and prostate cancer. Carcinogenesis (Lond) 18:641-644.
- Hayashi S, Watanabe J, Kawajiri K. 1992. High susceptibility to lung cancer analyzed in terms of combined genotypes of P4501A1 and Mu-class glutathione S-transferase genes. Japanese Journal of Cancer Research 83:866-870.
- Hayes JD, Pulford DJ. 1995. The glutathione S-transferase supergene family: regulation of GST and contribution of the isoenzymes to cancer chemoprotection and drug resistance. Critical Reviews in Biochemistry & Molecular Biology 30:445-600.
- Hirvonen A. 1999. Polymorphism of xenobiotic-metabolizing enzymes and susceptibility to cancer. Environmental Health Perspectives 107:37-47.
- Johansson AS, Stenberg G, Widersten M, Mannervik B. 1998. Structure activity relationships and thermal stability if human glutathione transferase P1-1 governed by H-site residue 105. Journal of Molecular
- Juronen E, Tasa G, Uuskual M, Pooga M, Mikelsaar AV. 1996. Purification, characterization and tissue distribution of human class theta glutathione 5-transferase T1-1. Biochemistry & Molecular Biology International 39:21-29.
- Kato S, Browman ED, Harrigan AM, Blomke B, Shields PG. 1995. Human lung carcinogen- DNA adduct levels mediated by genetic polymorphisms in vivo. Journal of National Cancer Institute 87:902-907.
- Kihara M, Kihara M, Noda K. 1999. Lung cancer risk of the GSTM1 null genotype is enhanced in the presence of the GSTP1 mutated genotype in male Japanese smokers. Cancer Letters 137:53-60.
- Kihara M, Noda K, Kihara M. 1995. Distribution of GSTM1 null genotype in relation to gender, age and smoking status in Japanese lung cancer patients. Pharmacogenetics 5:574-579.
- Kiyohara C, Yamamura KI, Nakanishi Y, Takayama K, Hara N. 2000. Polymorphisms in GSTM1, GSTT1 and GSTP1 and susceptibility to lung cancer in a Japanese population. Asian Pacific Journal of Cancer Prevention 1:293-298.
- Landi S. 2000. Mammalian class theta GST and differential susceptibility to carcinogens: a review. Mutation Research 463:247-283.
- LeMarchand L, Sivaraman L, Pierce A, Seifried A, Lum A, Wilkens LR, Lau AF. 1998. Association of CYP1A1, GSTM1 and CYP2E1 polymorphisms with lung cancer suggest cell type specificities to tobacco carcinogens. Cancer Research 58:4858-4863.
- Lewis SJ, Cherry NM, Niven RMcl, Barber PV, Povey AC. 2002. GSTM1, GSTT1 and GSTP1 polymorphisms and lung cancer risk. Cancer Letters 180:165-171.
- Mace K, Bowman ED, Vautravers P, Shields PG, Harris CC, Pfeifer AM. 1998. Characterisation of xenobiotic-metabolising enzyme expression in human bronchial mucosa and peripheral lung tissues. European Journal of Cancer 34:914-920.
- Mainwaring GW, Williams SM, Foster JR, Tugwood J, Green T. 1996. The distribution of theta class GST in the liver and lung of mouse, rat and human. Biochemical Journal 318:297-303.
- Mannervik B. 1985. The isoenzymes of glutathione transferases. Advances in Enzymology & Related Areas of Molecular Biology 57:357-417.



- Miller DP, Liu G, DeVivo I, Lynch TJ, Wain JC, Su L, Christianias DC. 2002. Combinations of the variant genotypes of GSTP1, GSTM1 and p53 are associated with an increased lung cancer risk. Cancer Research 62:2819-2823.
- Mironova NJ, Wikman H, Bouchardy C, Voho A, Dayer P, Benhamou S, Hirvonen A. 1998. Role of glutathione S-transferase GSTM1, GSTM3, GSTP1 and GSTT1 genotypes in modulating susceptibility to smoking-related lung cancer. Pharmacogenetics 8:495-502.
- Mishra DK, Kumar A, Srivastava DS, Mittal RD. 2004. Allelic variation of GSTT1, GSTM1 and GSTP1 genes in north Indian population. Asian Pacific Journal of Cancer Prevention 5:362-365.
- Mittal RD, Srivastava DS, AM, BM. 2005. Genetic polymorphism of drug metabolizing enzymes (CYP2E1, GSTP1) and susceptibility to bladder cancer in North India. Asian Pacific Journal of Cancer Prevention 6:6-9.
- Mittal RD, Srivastava DS, Kumar A, Mittal B. 2004. Polymorphism of GSTM1 and GSTT1 genes in prostate cancer: a study from north India, Indian Journal of Cancer 41:115-119.
- Naveen AT, Adithan C, Padmaja N, Shashinandran CH, Abraham BK, Satyanarayana K, Anitha P, Gerard N, Krishnamoorthy R. 2004. Glutathione-S-transferase M1 and T1 null genotypes distribution in south Indians. European Journal of Clinical Pharmacology 60:403-406.
- Pass HI, Mitchell JB, Johnson DH, Turrisi AT. 1996. Lung cancer: Principles and Practice. Philadelphia, PA: Lippincott-Raven Publishers. p. 697-710.
- Pemble S, Schroeder KR, Spencer SR, Meyer DJ, Hallier E, Bolt HM, Ketterer B, Taylor JB. 1994. Human gluatathione S-transferase Theta (GSTT1): cDNA cloning and the characterization of a genetic polymorphism. Biochemical Journal 300:271-276.
- Rebbeck TR. 1997. Molecular epidemiology of the human glutathione S-transferase genotypes GSTM1 and GSTT1 in cancer susceptibility. Cancer Epidemiology Biomarkers & Prevention 6:733–743.
- Ryberg D, Skaug V, Hewer A, Phillips DH, Harries LW, Wolf CR, Ogreid A, Ulvik P, Vu P, Haugen A. 1997. Genotypes of glutathione transferase M1 and P1 and their significance for lung DNA adduct levels and cancer risk. Carcinogenesis 18:1285-1289.
- Seideigard J, Pero RW, Markowitz MM, Roush G, Miller DG, Beattie EJ. 1990. Isoenzyme(s) of glutathione transferase (class Mu) as marker for the susceptibility to lung cancer: a follow up study. Carcinogenesis
- Sharma A, Sharma JK, Murthy NS, Mitra NB. 2004. Polymorphisms at GSTM1 and GSTT1 gene loci and susceptibility to cervical cancer in Indian population. Neoplasma 51:12-16.
- Skuladottir H, Autrup H, Autrup J, Tjoenneland A, Overvad K, Ryberg D, Haugen A, Olsen JH. 2005. Polymorphisms in genes involved in xenobiotic metabolism and lung cancer risk under the age of 60 years. A pooled study of lung cancer patients in Denmark and Norway. Lung Cancer 48:187-199.
- Sobti RC, Sharma S, Joshi A, Jindal SK, Janmeja A. 2004. Genetic polymorphisms of CYP1A1, CYP2E1, GSTM1 and GSTT1 genes and lung cancer susceptibility in north Indian population. Molecular & Cellular Biochemistry 266:1-9.
- Sreelekha TT, Tamadas K, Pandey M, Thomas G, Nalinakumari KR, Pillai MR. 2001. Genetic polymorphism of CYP1A1, GSTM1 and GSTT1 genes in Indian oral cancer. Oral Oncology 37:593-598.
- Srivastava DS, Kumar A, Mittal B, Mittal RD. 2004. Polymorphism of GSTM1 and GSTT1 genes in bladder cancer: a study from north India. Archives of Toxicology 78:430-434.
- Srivastava DS, Mandhani A, Mittal B, Mittal RD. 2005. Genetic polymorphism of glutathione-S-transferase gene (GSTM1, GSTT1 and GSTP1) and susceptibility to prostate cancer in Northern India. BJU International 95:170-173.
- Strange RC, Ellison T, Ichii-Jones F, Bath J, Hoban P, Lear JT, Smith AG, Hutchinson PE, Osborne J, Bowers B, Jones PW, Fryer AA. 1999. Cytochrome P450 CYP2D6 genotypes: association with hair colour, Breslow thickness and melanocyte stimulating hormone receptor alleles in patients with malignant melanoma. Pharmacogenetics 9:269-276.
- Stucker I, Hirvonen A, De Waziers I, Cabelguenne A, Mitrunen K, Cenee S, Koum-Besson E, Hemon D, Beaune P, Loriot MA. 2002. Genetic polymorphisms of glutathione S-transferases as modulators of lung cancer susceptibility. Carcinogenesis 23:1475-1481.
- Sundberg K, Johansson AS, Stenberg G, Widersten M, Seidel A, Mannervik B, Jernstrom B. 1998. Differences in the catalytic efficiencies of allelic variants of glutathione transferase P1-1 towards carcinogenic diol epoxides of polycyclic aromatic hydrocarbons. Carcinogenesis 19:433-436.
- Terrier P, Townsend AJ, Coindre JM, Triche TJ, Cowan KH. 1990. An immunohistochemical study of pi class glutathione S-transferase expression in normal human tissue. American Journal of Pathology 137:845-853.



- To-Figueras J, Gene M, Gomez-Catalan J, Pique E, Borrego N, Corbella J. 2001. Lung cancer susceptibility in relation to combined polymorphisms of microsomal epoxide hydrolase and glutathione S-transferase P1. Cancer Letters 173:155-162.
- Vos RME, Blanderen PJV. 1990. Glutathione S-transferases in relation to their role in the biotransformation of xenobiotics. Chemico-Biol Interactions 75:241-265.
- Wang J, Deng Y, Cheng J, Ding J, Tokudome S. 2003. GST genetic polymorphisms and lung adenocarcinoma susceptibility in a Chinese population. Cancer Letters 201:185-193.
- Watson MA, Stewart RK, Smith BJ, Massey TE, Bell DA. 1998. Human glutathione S-transferase P1 polymorphisms: relationship to lung tissue enzyme activity and population frequency distribution. Carcinogenesis (Lond) 19:275-280.
- Zheng Y, Song H, Higgins JPT, Pharoah P, Danesh J. 2006. Five glutathione S-transferase gene variants in 23 452 cases of lung cancer and 30 397 controls: meta-analysis of 130 studies. PLoS Medicine 3: 0524-0534.

